

Chromosomal Inversion Polymorphism Leads to Extensive Genetic Structure: A Multilocus Survey in *Drosophila subobscura*

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Manuscript received June 23, 2004

Accepted for publication December 13, 2004

ABSTRACT

The adaptive character of inversion polymorphism in *Drosophila subobscura* is well established. The O_{ST} and O_{3+4} chromosomal arrangements of this species differ by two overlapping inversions that arose independently on O_3 chromosomes. Nucleotide variation in eight gene regions distributed along inversion O_3 was analyzed in 14 O_{ST} and 14 O_{3+4} lines. Levels of variation within arrangements were quite similar along the inversion. In addition, we detected (i) extensive genetic differentiation between arrangements in all regions, regardless of their distance to the inversion breakpoints; (ii) strong association between nucleotide variants and chromosomal arrangements; and (iii) high levels of linkage disequilibrium in intralocus and also in interlocus comparisons, extending over distances as great as ~ 4 Mb. These results are not consistent with the higher genetic exchange between chromosomal arrangements expected in the central part of an inversion from double-crossover events. Hence, double crossovers were not produced or, alternatively, recombinant chromosomes were eliminated by natural selection to maintain coadapted gene complexes. If the strong genetic differentiation detected along O_3 extends to other inversions, nucleotide variation would be highly structured not only in *D. subobscura*, but also in the genome of other species with a rich chromosomal polymorphism.

CHROMOSOMAL inversion polymorphism is a common feature of the genome in the *Drosophila* genus. About 60% of *Drosophila* species are polymorphic for paracentric inversions in natural populations (POWELL 1997). The geographic distribution of inversions in many species and the seasonal change in frequency detected in some species strongly support that chromosomal polymorphism is adaptive (DOBZHANSKY 1970; KRIMBAS 1992; LEVITAN 1992; and others). Moreover, the reduced recombination in inversion heterokaryotypes (STURTEVANT 1926) led to the proposal that inversions could maintain complexes of coadapted linked genes favored by natural selection under particular conditions. Overdominance, frequency-dependent selection, or variable selection in time or space can contribute to the adaptive character of chromosomal polymorphism (see KRIMBAS and POWELL 1992; POWELL 1997).

For an advantageous inversion, the action of directional selection would rapidly drive the new arrangement to its equilibrium frequency. As a result of this rapid increase, all regions included in the new arrange-

ment would be completely depleted of variation even when the inversion had reached a relatively high frequency. Indeed, inverted chromosomes would initially be monomorphic for the particular haplotype captured by the inversion, which would include not only members of the coadapted gene complex but also neutral variants. The establishment of an inversion can thus be envisaged as a partial hitchhiking or selective sweep (MAYNARD SMITH and HAIGH 1974) that would lead to an initial genetic differentiation of inverted and noninverted chromosomes. Moreover, new mutations arising independently in the different arrangements would contribute to their further differentiation. Genetic exchange between chromosomal arrangements, either by gene conversion or by double crossover, could, however, erode any genetic differentiation. Most important, it could break down the coadapted gene complexes putatively underlying the selective advantage of inversions.

In the absence of selection, genetic differentiation would decay according to the rate of genetic exchange among arrangements. The gene conversion rate would be uniformly distributed along the inversion loop, whereas the contribution of double crossovers to genetic exchange would be considerably higher in the central part of the inversion loop (NAVARRO *et al.* 1997). Under this scenario (*i.e.*, in which genetic exchange increases with physical distance to inversion breakpoints), genetic differentiation among arrangements would be weaker in the central part of the loop than near the breakpoints (NAVARRO *et al.* 2000). In contrast, if selection were

Sequence data from this article have been deposited with the EMBL/GenBank Data Libraries under accession nos. AJ849711–AJ849884.

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maintaining coadapted gene complexes, it would counteract the homogenizing effect of genetic exchange on members of the complex. The differential action of selection would cause different levels of genetic differentiation along the inversion, but no relationship would be expected between the level of differentiation and the physical distance to breakpoints. Analysis of nucleotide variation along an inversion can thus inform us about the role played by natural selection in the establishment and maintenance of chromosomal polymorphism.

The O_{3+4}/O_{ST} system of *Drosophila subobscura* presents several distinctive features that make it especially suitable to detect the action of selection on chromosomal polymorphism through the study of nucleotide variation. First, the O_{3+4} and O_{ST} chromosomal arrangements differ by two overlapping inversions (inversions 3 and 4) that arose independently on the ancestral O_3 arrangement (RAMOS-ONSINS *et al.* 1998), which is now extinct in *D. subobscura* (Figure 1). This independent origin (which could be regarded as sampling a single O_3 chromosome twice) would result in an initial lack of nucleotide variation within an arrangement and in the initial presence of fixed differences between arrangements. Second, the existence of parallel latitudinal clines for these arrangements, both in Europe (KRIMBAS 1992) and in the recently colonized areas of North and South America, would support their adaptive character (PREVOSTI *et al.* 1988). Third, the $O_{3+4}-O_3-O_{ST}$ complex would conform to the Wallace rule of triads for partially overlapping inversions (WALLACE 1953; KRIMBAS 1992). According to this rule, elimination of the central member of a chromosomal triad would contribute to more efficiently maintaining longer coadapted gene complexes, since genetic exchange would be greatly reduced between the two external arrangements. Fourth, there is evidence of strong genetic differentiation between O_{ST} and O_{3+4} at loci near the distal breakpoint of inversion O_3 (ROZAS and AGUADÉ 1993, 1994; NAVARRO-SABATÉ *et al.* 1999). And fifth, the rather old age of O_{ST} and O_{3+4} (ROZAS and AGUADÉ 1994) suggests that recombination may have eroded the initial association between nucleotide variants and chromosomal arrangements. These features, and particularly the derived character of both arrangements and their age, differentiate the O_{ST}/O_{3+4} inversion system from others where variation at multiple regions has been surveyed (HASSON and EANES 1996; LAAYOUNI *et al.* 2003; MOUSSET *et al.* 2003; SCHAEFFER *et al.* 2003).

Here, we have analyzed the level and pattern of nucleotide variation in eight gene regions in a sample of O_{3+4} and O_{ST} chromosomes collected from a single natural population. These regions differ in their physical distance to the O_3 inversion breakpoints and completely cover this inversion. Our results show that genetic differentiation between O_{ST} and O_{3+4} chromosomes is strong and extends homogeneously all over the inversion. Therefore, genetic exchange between arrangements has been

strongly suppressed even in the central part of the inversion loop. The strong differentiation detected might be explained either by the absence of double crossovers in the O_3 inversion loop or by the elimination of double-crossover products by natural selection. The maintenance of the O_{3+4} and O_{ST} arrangements in natural populations of *D. subobscura* would have caused genetic variation at loci associated with these arrangements to be strongly structured.

MATERIALS AND METHODS

Isolation of genomic regions: Recombinant phages were isolated from the IPP246 genomic library of *D. subobscura* and amplified following standard procedures (SAMBROOK *et al.* 1989). Phage DNA was purified with the QIAGEN (Chatsworth, CA) lambda mini kit following manufacturer's instructions. DNA was labeled with 16-bio-dUTP and *in situ* hybridized on polytene chromosomes of *D. subobscura* according to SEGARRA and AGUADÉ (1992). A homokaryotypic O_{3+4} strain (*ch cu*) and an isochromosomal O_{ST} line were used for this purpose. Probes were mapped on the *D. subobscura* cytological map (KUNZE-MÜHL and MÜLLER 1958).

Phage DNA was digested with suitable restriction enzymes to release the arms, cloned into pBluescript SK+, and subsequently used to transform XL1-Blue *Escherichia coli* competent cells (Stratagene, La Jolla, CA). Insert sizes of recombinant plasmids were screened by PCR (KILGER and SCHMID 1994). DNA from plasmids with differing insert sizes was purified and both ends of each insert were sequenced. Inserts were completely sequenced by primer walking only in those cases where partial sequences showed high similarity to known or predicted genes of the 3R chromosomal arm of *Drosophila melanogaster* (which is homologous to the *D. subobscura* O chromosome).

Fly samples: Twenty-eight isochromosomal lines for the O chromosome established from a natural population of *D. subobscura* (ROZAS and AGUADÉ 1994; NAVARRO-SABATÉ *et al.* 1999) were used in this study: 14 O_{ST} and 14 O_{3+4} lines. A highly inbred *Drosophila madeirensis* line was also used for interspecific comparisons.

DNA sequencing: Genomic DNA from frozen flies was extracted using the DNA tissue kit (QIAGEN) following manufacturer's instructions, and the selected regions were subsequently PCR amplified using 21-mer primers. PCR conditions and amplification primers for the six newly reported regions are available in supplementary Figure 1 at <http://genetics.org/supplemental/>. Sequencing reactions were carried out with the ABI Prism BigDye Terminators 3.0 cycle sequencing kit (Applied Biosystems, Foster City, CA). Partial sequences were assembled with the SeqEd 1.03 program (HAGEMANN and KWAN 1997). Complete sequences were multiply aligned with the Clustal W program (THOMPSON *et al.* 1994) and further edited with the BioEdit 5.0.2 program (HALL 1999).

Data analysis: Analyses were based on the DNA sequences from the six newly reported regions and on the sequences from the *AcpH-1* (NAVARRO-SABATÉ *et al.* 1999; EMBL accession nos. AJ389424-AJ389476 and Y18840) and *rp49* (ROZAS and AGUADÉ 1994; RAMOS-ONSINS *et al.* 1998; accession nos. X80076-X80109 and Y09708) gene regions of *D. subobscura* and *D. madeirensis*. Analyses were performed for each region separately and for a single concatenated data set comprising those gene regions sequenced in the same 28 lines (*i.e.*, the six newly studied regions and *AcpH-1*).

Standard parameters of nucleotide polymorphism were esti-

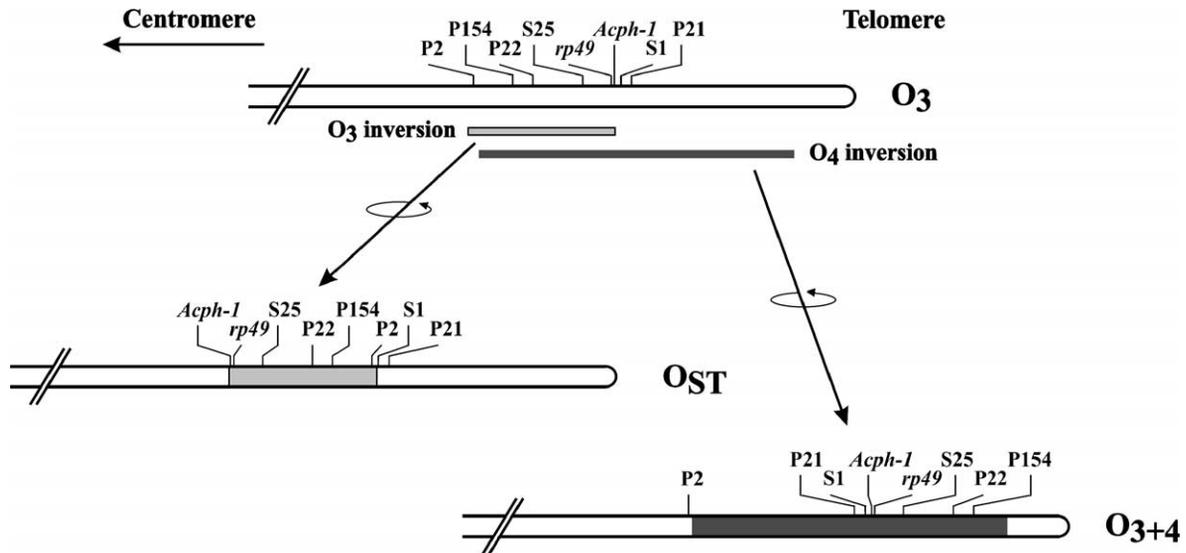


FIGURE 1.—Location of the eight gene regions in different chromosomal arrangements of the *D. subobscura* O chromosome. Their distribution along the ancestral O_3 arrangement is shown at the top. The effect of the O_3 (shading) and O_4 (solid) inversions on the location of the regions studied is also shown.

mated: the number of segregating sites in the sample (S), the minimum number of mutations (η), nucleotide diversity (π ; NEI 1987), and heterozygosity per site (θ ; WATTERSON 1975). The nucleotide divergence per silent site (K_{sil}) was estimated according to NEI and GOJOBORI (1986). The level of genetic differentiation between arrangements was estimated as D_{XY} (NEI 1987) and F_{ST} (HUDSON *et al.* 1992a) and its significance established using the K^* test statistic (HUDSON *et al.* 1992b). Gene conversion tracts were detected following BETRÁN *et al.* (1997). The probability that the observed number of polymorphisms shared between arrangements was due to recurrent mutation was estimated from the hypergeometric distribution as described in ROZAS and AGUADÉ (1994). The recombination length of the O_3 inversion was obtained considering a total length of 228.3 cM for the O chromosome of *D. subobscura* (LOUKAS *et al.* 1979). The physical distance between regions (or between a region and the nearest breakpoint) was estimated assuming that the euchromatic portion of the *D. subobscura* genome has 120 Mb (ADAMS *et al.* 2000) that are homogeneously distributed.

Linkage disequilibrium (LD) between pairs of parsimony informative sites (and association between informative sites and chromosomal arrangement) was estimated by the r^2 statistic (HILL and ROBERTSON 1968), and its statistical significance assessed by the χ^2 test with Bonferroni's correction for multiple comparisons (WEIR 1996). The overall level of LD was measured as ZnS (KELLY 1997) for parsimony informative sites (ZnS_i).

Neutrality tests (HUDSON *et al.* 1987; TAJIMA 1989; FU and LI 1993) were performed separately for the O_{ST} and O_{3+4} samples. Multilocus tests could also be performed within a chromosomal arrangement, given that recombination between regions was high and, therefore, that these regions have independent evolutionary histories. Statistical significance for all tests was assessed by coalescent simulations (10,000 independent replicates) conditioned on S under the conservative assumption of no intragenic recombination. *D. madeirensis* was used as the outgroup in those tests that required interspecific data. The DnaSP program 4.0 (ROZAS *et al.* 2003) was used to perform most of the analyses, and the HKA program (HEY 2004) for the multilocus tests.

Gene genealogies were reconstructed by the neighbor-join-

ing method (SAITOU and NEI 1987) as implemented in the MEGA 2.1 program (KUMAR *et al.* 2001). Genetic distances were obtained according to JUKES and CANTOR (1969). Bootstrap values were obtained after 1000 replicates.

RESULTS

Isolation of gene regions: A total of 200 recombinant phages randomly isolated from a *D. subobscura* genomic library were used for *in situ* hybridization on polytene chromosomes of this species. Of the ~ 100 phages that gave a unique signal, 34 mapped on the O chromosome. Of the 11 recombinant phages that hybridized in or around the O_3 inversion, 6 (S25, P22, P154, P2, S1, and P21) exhibited sequence similarity to genes on the 3R chromosomal arm of *D. melanogaster* (see supplementary Table 1 at <http://genetics.org/supplemental/> for relevant information about these regions). The location of the six isolated regions, plus that of *rp49* and *Acph-1*, along the O chromosome of *D. subobscura* is shown in Figure 1.

Nucleotide polymorphism and genetic differentiation between arrangements: The multiple alignment of the six newly reported gene regions in the 28 lines of *D. subobscura* consisted of 11,542 sites after excluding sites with alignment gaps. A total of 600 nucleotide polymorphic sites (293 singletons), which correspond to at least 612 mutations, were detected: 173 in coding regions (52 nonsynonymous and 121 synonymous) and 439 in noncoding regions (see supplementary Figure 2 at <http://genetics.org/supplemental/>). A summary of nucleotide variation in each region is shown in supplementary Table 2 at <http://genetics.org/supplemental/>. Some indel polymorphisms were also detected, mainly in noncoding regions.

TABLE 1
Genetic differentiation and gene flow between the O_{ST} and O_{3+4} chromosomal arrangements

	<i>AcpH-1</i>	<i>rp49</i>	S25	P22	P154	P2	S1	P21	Concatenated data ^a
Shared	19	12	22	6	4	2	6	3	62
Fixed	3	2	0	9	10	3	5	4	34
S_{X1}	73	34	36	32	39	50	12	27	269
S_{X2}	72	44	73	64	52	69	28	56	414
D_{XY}	0.0191	0.0145	0.0143	0.0170	0.0157	0.0157	0.0138	0.0106	0.0153
F_{ST}	0.43	0.50	0.44	0.52	0.66	0.54	0.59	0.51	0.52
K_S^*	3.15***	2.39***	2.78***	2.59***	2.51***	2.7***	1.92***	2.49***	4.60***

Shared, polymorphic sites segregating for the same two variants in both arrangements; Fixed, fixed differences among arrangements; S_{X1} , sites polymorphic in O_{ST} and monomorphic in O_{3+4} ; S_{X2} , sites polymorphic in O_{3+4} and monomorphic in O_{ST} ; D_{XY} , average number of nucleotide differences per site between arrangements; K_S^* , genetic differentiation test statistic (see MATERIALS AND METHODS); F_{ST} , proportion of nucleotide diversity attributable to variation among arrangements. *** $P < 0.001$.

^a Analysis performed in the concatenated data set that includes all six newly reported regions and only 28 of the 41 *AcpH-1* sequences (see MATERIALS AND METHODS).

Estimates of genetic differentiation between the O_{ST} and O_{3+4} arrangements were quite similar for the different regions (Table 1). Genetic differentiation was strong in each region as well as in the concatenated data set. Despite the significant genetic differentiation, all regions presented shared polymorphisms that in only three cases (P154, P2, and P21) could be explained by recurrent mutation. Genetic exchange between arrangements would therefore be necessary to explain the observed number of shared polymorphisms detected in *AcpH-1*, *rp49*, S25, P22, and S1. Indeed, genetic exchange could have contributed to the shared polymorphisms in all regions, since gene conversion tracts were identified in all but two regions (S1 and P21). No relationship (Figure 2a) was detected between the level of genetic differentiation and the distance to the nearest breakpoint (Kendall's $\tau = 0.143$, $P = 0.310$; Spearman's $\rho = 0.167$, $P = 0.347$).

Nucleotide variation estimates (Table 2) were obtained separately for each chromosomal arrangement, given the strong genetic differentiation detected. Estimates of nucleotide diversity (π_{total} and π_{sil}) were higher in O_{3+4} than in O_{ST} for all regions but P154, where they were similar in both arrangements. No relationship was detected, in either O_{3+4} or O_{ST} , between levels of silent nucleotide diversity within a chromosomal arrangement and physical distance to the nearest inversion breakpoint (Figure 3). Regions close to breakpoints did not show any reduction in nucleotide diversity. In fact, *AcpH-1* shows the highest π_{sil} value, in both O_{ST} and O_{3+4} , despite its tight linkage to the proximal breakpoint of the O_3 inversion. However, *AcpH-1* also showed the highest K_{sil} estimates, suggesting that this gene has a high neutral mutation rate. The direct relationship expected under the neutral model between levels of silent polymorphism and divergence (Table 2) was contrasted by the HKA test (HUDSON *et al.* 1987) using *D. madeirensis* as the outgroup. None of the tests performed between

pairs of gene regions yielded a significant result in either O_{ST} or O_{3+4} . A similar result was obtained in the multilocus test performed within arrangements (for O_{ST} , $\chi^2 = 1.87$, 7 d.f., $P = 0.96$; for O_{3+4} , $\chi^2 = 0.84$, 7 d.f., $P = 0.99$). Therefore, there is no significant heterogeneity in the ratio of polymorphism to divergence among the different regions.

Linkage disequilibrium analysis: Association between chromosomal arrangements (O_{ST} and O_{3+4}) and the variants present at informative polymorphic sites was analyzed (see supplementary Figure 3 at <http://genetics.org/supplemental/>). A total of 228 of the 385 informative sites in the concatenated data set (59.15%) showed a significant association ($P < 0.05$) with chromosomal arrangements. The association remained significant after Bonferroni correction in 34 sites (8.8%), which correspond to fixed differences between arrangements. A similar result was obtained for 48 informative sites in the *rp49* data set (43.7% and 20.8% of significant associations prior and after Bonferroni correction, respectively). No relationship was detected between the level of the association in each region (measured as the average r^2 value) and distance to the nearest breakpoint (Figure 2b).

The detected associations between variants at nucleotide sites and chromosomal arrangement should result in linkage disequilibrium between polymorphic nucleotide sites themselves. LD in the concatenated data set was analyzed first including all sequences (total sample) and then separately for O_{3+4} and O_{ST} . In the concatenated total data set with 385 informative sites, 28.8% of the pairwise comparisons showed significant LD ($P < 0.05$; Table 3). This percentage dropped to $\sim 5\%$ when each chromosomal arrangement was analyzed separately. Global estimates of LD, measured as ZnD_i , were also higher in the total sample than within the chromosomal arrangement: 0.1330 in the total sample, 0.0839 in O_{ST} , and 0.0845 in O_{3+4} . Recombination in homo-

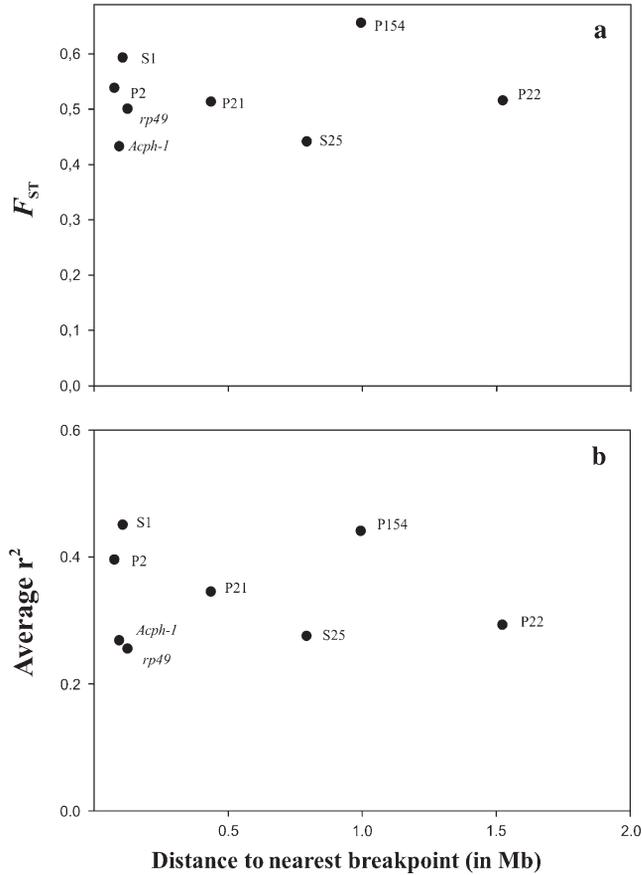


FIGURE 2.—(a) Genetic differentiation between chromosomal arrangements in gene regions distributed along the O_3 inversion. The distance between each region and the nearest O_3 inversion breakpoint was estimated assuming a homogeneous DNA distribution along the O_3 inversion (3.5 Mb). (b) Association between chromosomal arrangement and variants at informative polymorphic sites in each gene region plotted *vs.* distance to the nearest breakpoint. In all regions, except *rp49*, association was estimated for the same 28 lines (concatenated data set).

karyotypes would explain the lower percentage of significant LD within arrangement.

Pairwise comparisons were further classified as intra-

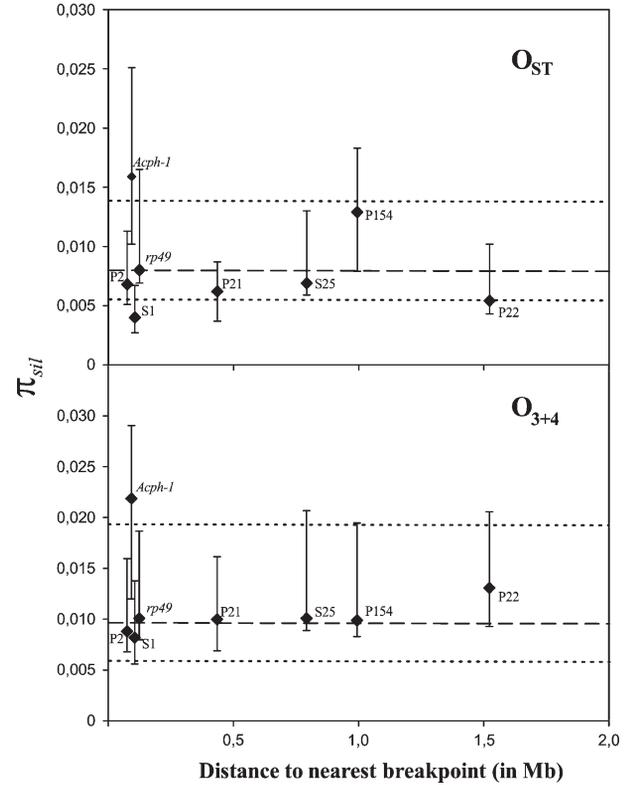


FIGURE 3.—Silent nucleotide diversity (π_{sil}) in the O_{ST} (top) and O_{3+4} (bottom) arrangements in each of the eight gene regions analyzed *vs.* distance to the nearest breakpoint of the O_3 inversion. π_{sil} estimates (diamonds) for each region and their 95% confidence intervals (vertical lines) are presented. Dashed and dotted horizontal lines show π_{sil} estimates within arrangements in the concatenated data set and the 95% confidence interval. Distance to the nearest breakpoint was inferred as in Figure 2.

locus and interlocus. The percentage of significant pairwise associations in the total sample was similar for intralocus (29.5%) and for interlocus (28.7%) comparisons, indicating that LD in the O_3 inversion extends over a long range. The presence of both arrangements therefore contributes to an increase in the level of intralocus

TABLE 2

Estimates of nucleotide polymorphism and divergence within chromosomal arrangements

	<i>Acph-1</i>		<i>rp49</i>		S25		P22		P154		P2		S1		P21	
	O_{ST}	O_{3+4}	O_{ST}	O_{3+4}	O_{ST}	O_{3+4}	O_{ST}	O_{3+4}	O_{ST}	O_{3+4}	O_{ST}	O_{3+4}	O_{ST}	O_{3+4}	O_{ST}	O_{3+4}
<i>n</i>	21	20	16	18	14	14	14	14	14	14	14	14	14	14	14	14
<i>S</i>	92	91	45	54	58	93	38	70	43	56	50	70	17	34	30	58
π_{total}	0.010	0.012	0.006	0.008	0.006	0.010	0.005	0.011	0.005	0.005	0.006	0.008	0.004	0.007	0.004	0.006
π_{sil}	0.016	0.022	0.008	0.010	0.007	0.010	0.005	0.013	0.013	0.010	0.007	0.009	0.004	0.008	0.006	0.010
θ_{sil}	0.018	0.021	0.012	0.014	0.009	0.015	0.007	0.015	0.013	0.014	0.009	0.012	0.005	0.010	0.006	0.012
K_{sil}	0.043	0.046	0.013	0.018	0.016	0.018	0.020	0.027	0.034	0.035	0.023	0.028	0.018	0.022	0.023	0.026

n, sample size or number of sequences; *S*, number of polymorphic sites; π_{total} , nucleotide diversity in all sites; π_{sil} , nucleotide diversity in silent sites; θ_{sil} , heterozygosity per silent site based on segregating sites; K_{sil} , divergence per silent site between *D. subobscura* and *D. madeirensis*.

TABLE 3

Linkage disequilibrium in the concatenated data set

Arrangements	Total (%)	Intralocus (%)	Interlocus (%)
Both arrangements ($n = 28$)	28.8	29.5	28.7
O_{ST}	5	10.8	3.9
O_{3+4}	5.3	11.1	3.6

Linkage disequilibrium measured as the percentage of significant ($P < 0.05$) pairwise comparisons by the χ^2 test.

LD and in the extent of interlocus disequilibrium. On the other hand, the level of LD was relatively reduced in both the intralocus and the interlocus analyses within arrangements (Table 3). This result can be explained again by recombination in homokaryotypes.

Global estimates of interlocus LD were also obtained for all pairwise comparisons between regions and compared with those for intralocus LD. As shown in Figure 4, ZnS_i estimates in O_{ST} and in O_{3+4} were higher for intralocus than for interlocus comparisons. In contrast, in the total sample, the intralocus and interlocus ZnS_i estimates were much more similar. Indeed, all interlocus estimates were within the range established by the intralocus estimates. Moreover, no relationship between interlocus ZnS_i estimates and the distance between pairs of regions was detected.

Pattern of polymorphism: Several statistical tests (Tajima 1989; Fu and Li 1993) were performed to assess whether the pattern of variation within arrangements conforms to expectations of the neutral equilibrium model of molecular evolution (see supplementary Table 3 at <http://genetics.org/supplemental/>). For individual regions, all test statistics were negative in O_{ST} and also in seven of the eight regions in O_{3+4} . This trend toward negative values was further analyzed using the multilocus test based on the mean value of Tajima's D statistic (\bar{D}). For both O_{ST} and O_{3+4} , the empirical D -value averaged across the eight regions studied was significantly lower (two-tailed test) than the average D -value obtained from the simulations: $\bar{D} (O_{3+4}) = -0.8666$, $P = 0.004$; $\bar{D} (O_{ST}) = -0.8349$, $P = 0.018$. A similar result was obtained for the multilocus test based on Fu and Li's D statistic (not shown). Therefore, an overall significant excess of low-frequency variants, mainly singletons, was detected in both arrangements.

Gene genealogy: Figure 5 shows the gene genealogy reconstructed from total variation in the concatenated total data set. Sequenced lines clearly cluster according to chromosomal arrangement, which is consistent with the strong genetic differentiation detected between arrangements. This clustering was supported by very high bootstrap percentages (100% for the O_{ST} and the O_{3+4} clusters) and was also detected when each region was analyzed separately, except S25 (in this region, an O_{3+4}

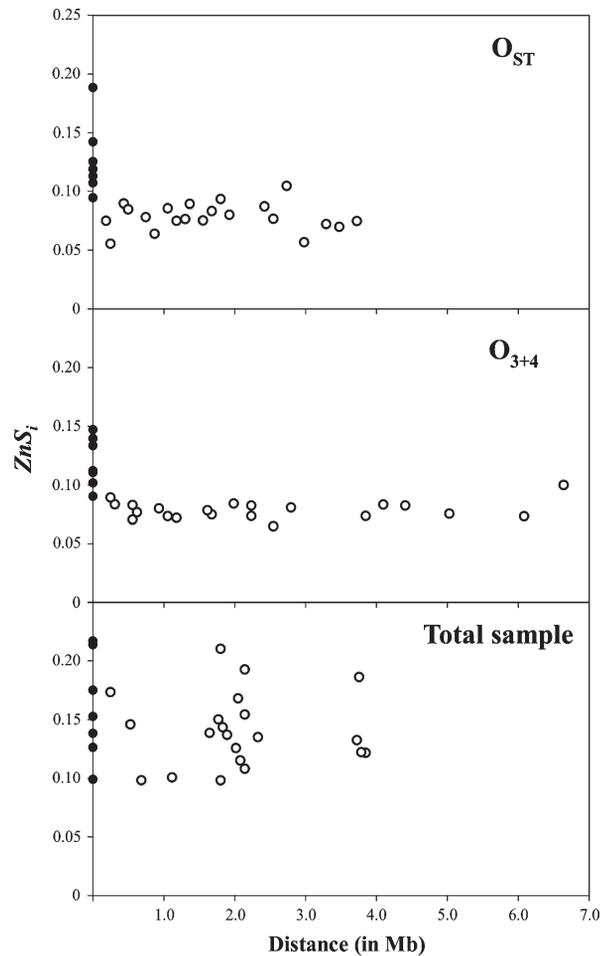


FIGURE 4.—Interlocus LD (open circles) in each chromosomal arrangement and in the total sample plotted *vs.* interlocus distance. Intralocus LD (solid circles) is also shown. Pairwise distances between regions are different in O_{ST} and O_{3+4} since these arrangements differ by two inversions (Figure 1); in the total sample, distances between regions were estimated as the average of the distances in O_{ST} and O_{3+4} .

line with a rather long gene conversion tract clustered with the O_{ST} lines). For each cluster, the genealogy is characterized by relatively short internal and long external branches, *i.e.*, a star-like genealogy.

DISCUSSION

The establishment and maintenance of inversion polymorphism in natural populations of *Drosophila* has been explained by a superior fitness of heterokaryotypes (DOBZHANSKY 1970). The pattern of variation detected in the present multilocus study is consistent with the action of natural selection in the establishment of O_{ST} and O_{3+4} . The general trend toward an excess of low-frequency variants in the derived arrangements O_{ST} and O_{3+4} , the significant multilocus neutrality tests, and the star-like genealogy within arrangements would reflect the partial hitchhiking or selective sweep that drove these arrangements to their equilibrium frequencies.

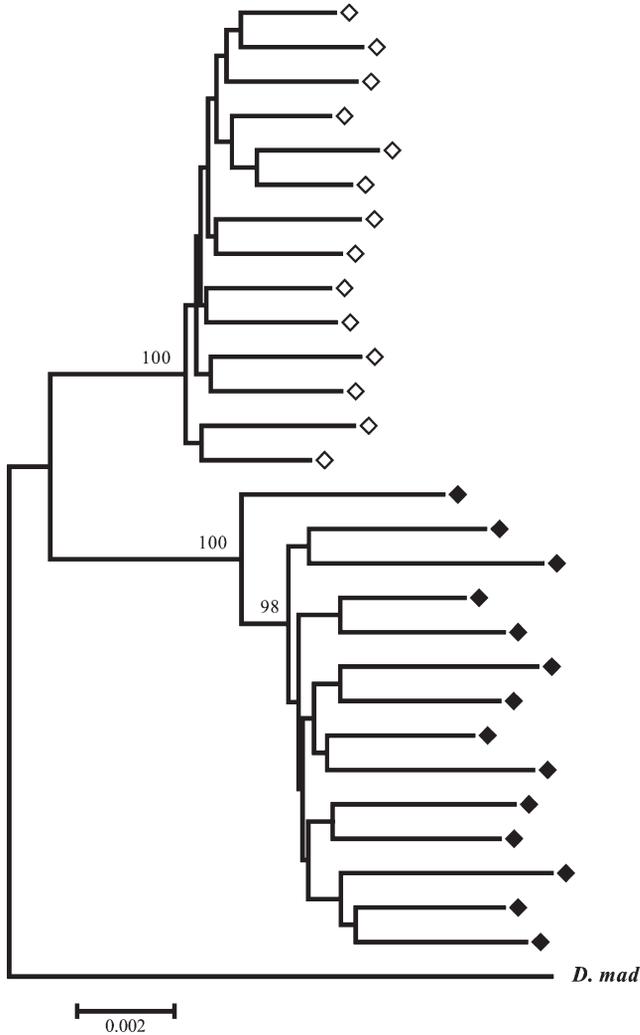


FIGURE 5.—Neighbor-joining gene genealogy based on total nucleotide variation in the concatenated data set (see MATERIALS AND METHODS). Bootstrap values $>95\%$ are shown. Open and solid diamonds refer to O_{ST} and O_{3+4} *D. subobscura* lines, respectively. *D. mad* refers to the *D. madeirensis* line used as outgroup.

After the partial selective sweep associated with the establishment of a new inversion, a strong depletion of variation is expected around the breakpoints and also in very close-by regions (ANDOLFATTO *et al.* 2001). Indeed, new variation in these regions can be introduced only by mutation, as gene conversion would be suppressed due to mechanical problems in synapses. Although some of the regions studied here are rather close to the breakpoints, none of them exhibits a reduction in variation. Indeed, estimates of π_{sil} in these regions are similar to, although slightly lower than, the value estimated for the *Acp70A* region of *D. subobscura* ($\pi_{sil} = 0.016$; CIRERA and AGUADÉ 1998), which is located in a chromosomal region not affected by inversions. Moreover, the polymorphism-to-divergence ratio is quite homogeneous among regions. These results, and the detection of gene conversion tracts in most of the regions

studied, indicate that their distance to the nearest breakpoint is high enough for gene conversion to have contributed to the recovery of variation.

The multilocus analysis reported here clearly indicates that genetic differentiation is strong and extends all over the inversion. Indeed, LD is as pervasive in interlocus as in intralocus comparisons, despite a 0.5–4 Mb range of interlocus distances (Figure 4). There is no evidence for the higher genetic exchange between arrangements expected in the central part of the inversion loop in the presence of gene conversion and double crossover (NAVARRO *et al.* 1997). The rather homogeneous distribution of genetic exchange detected across the inversion would indicate, therefore, that no double crossovers were produced in the inversion loop or, alternatively, that selection has acted against the recombinant chromosomes.

The occurrence over evolutionary time of double crossovers inside an inversion loop may be contingent on its length and age. Considering the empirical values of interference in *Drosophila*, NAVARRO *et al.* (1997) suggested that double crossover is unlikely only in short inversions (<20 cM). The estimated length of the O_3 inversion (27.4 cM) would thus *a priori* support that double crossovers could have contributed, at least partly, to the genetic exchange in this inversion. In addition, the time elapsed since its origin (0.25–0.3 MYA; ROZAS and AGUADÉ 1994) is long enough for double crossovers to have broken the initial associations, at least in the central part of the inversion loop. Double crossovers also have not been effective in eroding the genetic differentiation in the central part of the ~ 65 -cM-long inversion that differentiates the O_{3+4} and O_{3+4+8} arrangements (ROZAS *et al.* 1999; NAVARRO-SABATÉ *et al.* 2003).

Accepting the occurrence of double crossovers, selection acting against the products of genetic exchange between chromosomal arrangements, and more specifically against double-crossover products, would be the most plausible explanation for the strong genetic differentiation detected in the eight regions studied. Indeed, epistatic fitness interactions among genes within the inversion would result in the lower fitness of those among-arrangement recombinants that affected the coadapted complex. Sets of coadapted linked genes would be broken more likely by double crossover than by gene conversion, as the lengths of the segments affected by gene conversion are much shorter (HILLIKER *et al.* 1994; BETRÁN *et al.* 1997). Consequently, selection would have acted mostly against double-crossover products.

The eight regions studied, which were chosen at random with the sole restriction being to cover the O_3 inversion, exhibited a strong genetic differentiation. For epistatic selection to explain this result, the regions need not be the targets of selection themselves, but they should be tightly linked to genes of the coadapted complex. Our observation would imply a rather high number of target genes or, alternatively, fewer genes with

stronger effects. Indeed, the high level of interlocus LD detected in the total sample of O_{ST} and O_{3+4} chromosomes (Figure 4) indicates that the regions linked to each arrangement have followed independent evolutionary histories. Therefore, the effects of coadapted complexes on nucleotide variation and genetic differentiation would be large and, at least for the O_3 inversion, might affect the complete inverted fragment. In *D. pseudoobscura*, the pattern of nucleotide variation detected in gene regions associated with the third chromosome arrangements also supports that epistatic selection maintains chromosomal polymorphism (SCHAEFFER et al. 2003). However, in this species, unlike in *D. subobscura*, there was a general trend toward a reduction of linkage disequilibrium with distance.

The strong genetic differentiation detected all along the O_3 inversion is remarkable regardless of whether it is a consequence of the lack of double crossovers inside the inversion loop or of the action of epistatic selection. The presence of O_{ST} and O_{3+4} in natural populations would cause a strong structuring of nucleotide variation that extends along more than the ~ 3.5 Mb of the O_3 inversion, since regions outside but close to breakpoints exhibit the same pattern. *D. subobscura* harbors a very rich chromosomal polymorphism in all chromosomes of the complement (except the dot-like element), and all individuals in natural populations are likely heterozygous for chromosomal inversions. If the results found in O_3 hold for other inversions, nucleotide variation in a major part of the *D. subobscura* genome might indeed be highly structured. A similar although less general pattern might be expected in other species where chromosomal polymorphism is more restricted. Chromosomal polymorphism would thus result in the presence of different gene pools with independent evolutionary fates, which might have major evolutionary consequences.

We thank Gema Blasco for technical support. We also thank Serveis Científico-Tècnics, Universitat de Barcelona, for automated DNA sequencing facilities. This work was supported by grants PB97-0918 and BMC2001-2906 from Comisión Interdepartamental de Ciencia y Tecnología, Spain, and grants 1999SGR-25 and 2001SGR-00101 from Comissió Interdepartamental de Recerca i Innovació Tecnològica, Generalitat de Catalunya, Spain, to M.A.

LITERATURE CITED

- ADAMS, M. D., S. E. CELNIKER, R. A. HOLT, C. A. EVANS, J. D. GOCAYNE et al., 2000 The genome sequence of *Drosophila melanogaster*. *Science* **287**: 2185–2195.
- ANDOLFATTO, P., F. DEPAULIS and A. NAVARRO, 2001 Inversion polymorphisms and nucleotide variability in *Drosophila*. *Genet. Res.* **77**: 1–8.
- BETRÁN, E., J. ROZAS, A. NAVARRO and A. BARBADILLA, 1997 The estimation of the number and the length distribution of gene conversion tracts from population DNA sequence data. *Genetics* **146**: 89–99.
- CIRERA, S., and M. AGUADÉ, 1998 Molecular evolution of a duplication: the sex-peptide (*Acp70A*) gene region of *Drosophila subobscura* and *Drosophila madeirensis*. *Mol. Biol. Evol.* **15**: 988–996.
- DOBZHANSKY, TH., 1970 *Genetics of the Evolutionary Process*. Columbia University Press, New York.
- FU, Y.-X., and W.-H. LI, 1993 Statistical tests of neutrality of mutations. *Genetics* **133**: 693–709.
- HAGEMANN, T. L., and S. P. KWAN, 1997 SeqEd: manipulation of sequence data and chromatograms from the ABI DNA sequencer analysis files. *Methods Mol. Biol.* **70**: 55–63.
- HALL, T. A., 1999 BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symp. Ser.* **41**: 95–98.
- HASSON, E., and W. F. EANES, 1996 Contrasting histories of three gene regions associated with In(3L)Payne of *Drosophila melanogaster*. *Genetics* **144**: 1565–1575.
- HEY, J., 2004 <http://lifesci.rutgers.edu/~hey/Heylab/HeylabSoftware.htm>.
- HILL, W. G., and A. ROBERTSON, 1968 Linkage disequilibrium in finite populations. *Theor. Appl. Genet.* **38**: 226–231.
- HILLIKER, A. J., G. HARAUZ, A. G. REAUME, M. GRAY, S. H. CLARK et al., 1994 Meiotic gene conversion tract length distribution within the *rosy* locus of *Drosophila melanogaster*. *Genetics* **137**: 1019–1026.
- HUDSON, R. R., M. KREITMAN and M. AGUADÉ, 1987 A test of neutral molecular evolution based on nucleotide data. *Genetics* **116**: 153–159.
- HUDSON, R. R., M. SLATKIN and W. P. MADDISON, 1992a Estimation of levels of gene flow from DNA sequence data. *Genetics* **132**: 583–589.
- HUDSON, R. R., D. D. BOOS and N. L. KAPLAN, 1992b A statistical test for detecting geographic subdivision. *Mol. Biol. Evol.* **9**: 138–151.
- JUKES, T. H., and C. R. CANTOR, 1969 Evolution of protein molecules, pp. 21–120 in *Mammalian Protein Metabolism*, edited by H. M. MUNRO. Academic Press, New York.
- KELLY, J. K., 1997 A test of neutrality based on interlocus associations. *Genetics* **146**: 1197–1206.
- KILGER, C., and K. SCHMID, 1994 Rapid characterization of bacterial clones by microwave treatment and PCR. *Trends Genet.* **10**: 149.
- KRIMBAS, C. B., 1992 The inversion polymorphism of *Drosophila subobscura*, pp. 127–220 in *Drosophila Inversion Polymorphism*, edited by C. B. KRIMBAS and J. R. POWELL. CRC Press, Boca Raton, FL.
- KRIMBAS, C. B., and J. R. POWELL (EDITORS), 1992 *Drosophila Inversion Polymorphism*, pp. 1–52. CRC Press, Boca Raton, FL.
- KUMAR, S., K. TAMURA, I. B. JAKOBSEN and M. NEI, 2001 MEGA2: molecular evolutionary genetics analysis software. *Bioinformatics* **17**: 1244–1245.
- KUNZE-MÜHL, E., and E. MÜLLER, 1958 Weitere Untersuchungen über die chromosomale Struktur und natürlichen Strukturtypen von *D. subobscura*. *Chromosoma* **9**: 559–570.
- LAAYOUNI, H., E. HASSON, M. SANTOS and A. FONTDEVILA, 2003 The evolutionary history of *Drosophila buzzatii*. XXXV. Inversion polymorphism and nucleotide variability in different regions of the second chromosome. *Mol. Biol. Evol.* **20**: 931–944.
- LEVITAN, M., 1992 Chromosomal variation in *Drosophila robusta* Sturtevant, pp. 221–338 in *Drosophila Inversion Polymorphism*, edited by C. B. KRIMBAS and J. R. POWELL. CRC Press, Boca Raton, FL.
- LOUKAS, M., C. B. KRIMBAS and Y. VERGINI, 1979 The genetics of *Drosophila subobscura* populations. IX. Studies on linkage disequilibrium in four natural populations. *Genetics* **93**: 497–523.
- MAYNARD SMITH, J., and J. HAIGH, 1974 The hitch-hiking effect of a favourable gene. *Genet. Res.* **23**: 23–35.
- MOUSSET, S., L. BRAZIER, M. L. CARIU, F. CHARTOIS, F. DEPAULIS et al., 2003 Evidence of a high rate of selective sweeps in African *Drosophila melanogaster*. *Genetics* **163**: 599–609.
- NAVARRO, A., E. BETRÁN, A. BARBADILLA and A. RUIZ, 1997 Recombination and gene flux caused by gene conversion and crossing over in inversion heterokaryotypes. *Genetics* **146**: 695–709.
- NAVARRO, A., A. BARBADILLA and A. RUIZ, 2000 Effect of inversion polymorphism on the neutral nucleotide variability of linked chromosomal regions in *Drosophila*. *Genetics* **155**: 685–698.
- NAVARRO-SABATÉ, A., M. AGUADÉ and C. SEGARRA, 1999 The relationship between allozyme and chromosomal polymorphism inferred from nucleotide variation at the *Acp1-1* gene region of *Drosophila subobscura*. *Genetics* **153**: 871–889.
- NAVARRO-SABATÉ, A., M. AGUADÉ and C. SEGARRA, 2003 Excess of nonsynonymous polymorphism at *Acp1-1* in different gene arrangements of *Drosophila subobscura*. *Mol. Biol. Evol.* **20**: 1833–1843.
- NEI, M., 1987 *Molecular Evolutionary Genetics*. Columbia University Press, New York.
- NEI, M., and T. GOJOBORI, 1986 Simple methods for estimating the

- numbers of synonymous and nonsynonymous nucleotide substitutions. *Mol. Biol. Evol.* **3**: 418–426.
- POWELL, J. R., 1997 *Progress and Prospects in Evolutionary Biology: The Drosophila Model*. Oxford University Press, New York.
- PREVOSTI, A., G. RIBÓ, L. SERRA, M. AGUADÉ, J. BALANÀ *et al.*, 1988 Colonization of America by *Drosophila subobscura*: experiment in natural populations supporting the adaptative role of chromosomal-inversion polymorphism. *Proc. Natl. Acad. Sci. USA* **85**: 5597–5600.
- RAMOS-ONSINS, S., C. SEGARRA, J. ROZAS and M. AGUADÉ, 1998 Molecular and chromosomal phylogeny in the obscura group of *Drosophila* inferred from sequences of the *rp49* gene region. *Mol. Phylogenet. Evol.* **9**: 33–41.
- ROZAS, J., and M. AGUADÉ, 1993 Transfer of genetic information in the *rp49* region of *Drosophila subobscura* between different chromosomal gene arrangements. *Proc. Natl. Acad. Sci. USA* **90**: 8083–8087.
- ROZAS, J., and M. AGUADÉ, 1994 Gene conversion is involved in the transfer of genetic information between naturally occurring inversions of *Drosophila*. *Proc. Natl. Acad. Sci. USA* **91**: 11517–11521.
- ROZAS, J., C. SEGARRA, G. RIBÓ and M. AGUADÉ, 1999 Molecular population genetics of the *rp49* gene region in different chromosomal inversions of *Drosophila subobscura*. *Genetics* **151**: 189–202.
- ROZAS, J., J. C. SANCHEZ-DELBARRIO, X. MESSEGUER and R. ROZAS, 2003 DnaSP, DNA polymorphism analyses by the coalescent and other methods. *Bioinformatics* **19**: 2496–2497.
- SAITOU, N., and M. NEI, 1987 The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol. Biol. Evol.* **4**: 406–425.
- SAMBROOK, J., E. F. FRITSCH, and T. MANIATIS, 1989 *Molecular Cloning: A Laboratory Manual*. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
- SCHAEFFER, S. W., M. P. GOETTING-MINESKY, M. KOVACEVIC, J. R. PEOPLES, J. L. GRAYBILL *et al.*, 2003 Evolutionary genomics of inversions in *Drosophila pseudoobscura*: evidence for epistasis. *Proc. Natl. Acad. Sci. USA* **100**: 8319–8324.
- SEGARRA, C., and M. AGUADÉ, 1992 Molecular organization of the X chromosome in different species of the obscura group of *Drosophila*. *Genetics* **130**: 513–521.
- STURTEVANT, A. H., 1926 A crossover reducer in *Drosophila melanogaster* due to inversion of a section of the third chromosome. *Biol. Zentralbl.* **46**: 697–702.
- TAJIMA, F., 1989 Statistical method for testing the neutral mutation hypothesis by DNA polymorphism. *Genetics* **123**: 585–595.
- THOMPSON, J. D., D. G. HIGGINS and T. J. GIBSON, 1994 CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res.* **22**: 4673–4680.
- WALLACE, B., 1953 On coadaptation in *Drosophila*. *Am. Nat.* **87**: 343–358.
- WATTERSON, G. A., 1975 On the number of segregating sites in genetical models without recombination. *Theor. Popul. Biol.* **7**: 256–276.
- WEIR, B. S., 1996 *Genetic Data Analysis II*. Sinauer Associates, Sunderland, MA.

Communicating editor: M. VEUILLE

