**Detailed protocol for colony embedding:** The following is a modification of a method for embedding yeast colonies for light and electron microscopy (1). After incubation of approximately 300 colonies on agar medium for the indicated time, an isolated colony (1-2 mm in diameter) and a small amount of the underlying agar medium was removed and placed on a microscope slide. Several drops of 2% agar (42°C) were placed on a microscope slide, the colony was immediately placed on the agar and then several drops of agar were placed on top of the colony. The resulting agar block was trimmed with a razor blade, and placed in a 3.5 ml borosilicate screw-cap vial (Fisher 03-339-21B). All subsequent incubations and washes used 1.5 -2.0 ml and were performed in the same vial. First, colonies were fixed by incubating in 2% paraformaldehyde / 2% glutaraldehyde for 7 days at 4°C. Agar blocks were then washed on ice by incubating twice for 15 minutes with 0.5 ml of 0.15M sodium cacodylate (pH 7.2), and then twice more for 5 minutes with OS buffer (100 mM KH$_2$PO$_4$, 10 mM MgCl$_2$, pH 6.0). To allow electron microscopy of sections, 1% OsO$_4$ in OS was added to vials to cover the agar block and incubated on ice in a fume hood for 1 hr, then washed twice with OS buffer on ice, and incubated in OS overnight at 4°C. After washing two times with cold water, the blocks were washed sequentially with 25%, 50%, 75%, 95% and 100% (twice) ethanol for 10 minutes each, and the blocks left overnight at 4°C in 100% ethanol. Ethanol was removed, Spurr’s reagent (Electron Microscopy Sciences) added, vials rotated for 15 minutes, then allowed to stand for 30 minutes, and the Spurr’s treatment repeated four more times. After the final wash, Spurr’s reagent was added to cover agar blocks (1.5-2.0 ml) and incubated for four hours, then the Spurr’s was replaced, the vial rotated overnight, and this last step repeated again the next day. Finally, each agar block was placed in a mold with 0.2 ml of Spurr’s and incubated at 60°C for four hours, topped off with Spurr’s and incubated at 60°C overnight. Sections (0.5 µ) from the central region of the colony were collected in distilled H$_2$O, placed on a drop of water, and dried on a 52°C heat block. Sections were stained with 1.0% toluidine blue, 1% Sodium Borate for 5-15 sec., washed under a stream of water, dried, covered in mounting media (KPL), and examined by light microscopy.


**Detailed Protocol for Cryosections:** A square of agar surrounding a 2-3 mm colony was cut from an agar plate and submerged in OCT compound (VWR) in a 15 X 15 X 5 mm cryomold (VWR) and incubated at room temperature for 90 min. The cryomold was then submerged for 20 sec in an isopentane bath cooled by liquid nitrogen and sectioned at -20°C -- -25°C using a Microm HM 505E cryostat. 20 µ sections through the central region of the colony were covered with mounting media (KPL) and examined using Nomarski optics.